Phylogenetic relationships of African caecilians (Amphibia: Gymnophiona): insights from mitochondrial rRNA gene sequences

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Abstract.—Africa (excluding the Seychelles) has a diverse caecilian fauna, including the endemic family Scolecomorphidae and six endemic genera of the more cosmopolitan Caeciliidae. Previous molecular phylogenetic studies have not included any caecilians from the African mainland. Partial 12S and 16S mitochondrial gene sequences were obtained for two species of the endemic African Scolecomorphidae and five species and four genera of African caeciliids, aligned against previously reported sequences for 16 caecilian species, and analysed using parsimony, maximum likelihood, Bayesian and distance methods. Results are in agreement with traditional taxonomy in providing support for the monophyly of the African caecilian genera Boulengerula and Schistometopum, and for the Scolecomorphidae. They disagree in indicating that the Caeciliidae is paraphyletic with respect to the Scolecomorphidae. Although more data from morphology and/or molecules will be required to resolve details of the interrelationships of the African caecilian genera, the data provide strong support for at least two origins of caecilians in which the eye is reduced and covered with bone, and do not support the hypotheses that the caecilian assemblages of Africa, and of East and of West Africa are monophyletic.

Key words.—Amphibians, biogeography, evolution, eyes, phylogeny, vertebrates, viviparity.

Caecilians (Gymnophiona) are one of the three extant orders of amphibians. The caecilian fauna of Africa, taken as excluding the Seychelles, includes the endemic family Scolecomorphidae (six species in two genera) and six genera and 16 species of the more cosmopolitan Caeciliidae, which also has representatives in the Seychelles, India, and Central and South America. African caecilians make up approximately 13% and 25% of the recognised caecilian species and genera respectively, and thus constitute a substantial proportion of known gymnophionan diversity. Previous molecular phylogenetic analyses (Hedges et al. 1993; Gower et al. 2002; Wilkinson et al. 2002) have included, at most, only a single African caecilian, the insular caeciliid Schistometopum thomense. Apart from an unconfirmed report from Central Africa (Nussbaum & Pfrender 1998), this caeciliid is known only from Sao Thome in the Gulf of Guinea. Thus we have no molecular phylogenetic insight into the relationships of any mainland African caecilians. Of the six currently recognised caecilian families (Nussbaum & Wilkinson 1989) only the Scolecomorphidae remains unstudied with regards to molecular data.
Building on the foundations provided by Hedges et al. (1993), Wilkinson et al. (2002) used partial 12S and 16S SSU mt DNA sequence data to provide well supported resolution of the phylogenetic relationships of representatives of the three families of caecilians present in India, and suggested that expanding the sampling of African caecilians was a priority for caecilian molecular phylogenetics.

Here we report new 12S and 16S SSU rDNA partial sequences for seven species of African caecilians, including the first sequences for representatives of the Scolecomorphidae, and the first sequences for any East African caeciliid. At the generic level, our sampling of African caecilians is incomplete only in the omission of the monotypic caeciliids Sylvacaecilia from Ethiopia and Idiocranium from Cameroon, and of the West African scolecomorphid Crotophatrema (three species). The new sequences increase the diversity of caecilians for which these comparative mitochondrial sequence data are available, from 16 to 23 of the approximately 160 currently recognised caecilian species, and from 11 to 15 of the 33 genera. The new sequences allow the first molecular tests of the monophyly of Scolecomorphus and the Scolecomorphidae, of Boulengerula and of Schistometopum, and investigation of the relationships of the caeciliid assemblages of East and West Africa to each other and a range of non-African caecilians.

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**Materials and Methods**

Details of voucher specimens are presented in Table 1. Sequencing methods are as given in Wilkinson et al. (2002). Sequences have been deposited in GenBank (Benson et al. 1998) with accession numbers A7450612 - A7450625.

The newly determined sequences were added to an alignment of concatenated partial 12S and 16S caecilian sequences (Wilkinson et al. 2002) and the alignment adjusted manually. Regions in which positional homology could not be assessed with confidence due to length variation were excluded, except where length variation was concentrated in a minority of taxa. In the latter case, regions of uncertainty in the alignment were represented by replacing the sequence data for the minority of taxa with missing entries. This increases the available data for the remaining majority of taxa and reduces the amount of useful information that is discarded. Following Wilkinson et al. (2002), the sequence of the rhinatrematid caecilian Epicrionops marmoratus was designated as a single outgroup and used to root trees.

Parsimony, maximum likelihood (ML) and distance analyses were performed with PAUP* 4.0b10 (Swofford 1998). LogDet and Maximum Likelihood distance (MLD) analyses used the minimum evolution objective function. ML and MLD analyses used models...
of evolution selected by Modeltest (Posada & Crandall 1998) and the corresponding estimated proportion of invariant sites was used in the LogDet analyses. Alignment gaps were treated as missing data. Tree searches were heuristic with 100 (parsimony and distance analyses) or 10 (ML) random addition sequences and TBR branch swapping. A Bayesian analysis was performed using MrBayes 2.01 (Huelsenbeck & Ronquist 2001) using a general time reversible (GTR) model, with rate variation across sites modelled with a discrete gamma distribution (G) and proportion of invariant sites (I). The Metropolis coupled, Markov chain Monte Carlo analysis was run with four chains for 1,000,000 generations. Trees were sampled every 1000 generations, with the first 1000 generations discarded as “burn in”.

A parsimony PTP test (Faith & Cranston 1991) was used to test the null hypothesis that the alignment has no more hierarchical structure than expected by chance alone (99 random permutations). Support for clades was measured with bootstrap proportions (Felsenstein 1985) (100 pseudoreplicates) and Bayesian posterior probabilities. Leaf stabilities based on the bootstrap difference measure (Thorley & Wilkinson 1999) were determined using RadCon (Thorley & Page 2000) from sets of bootstrap trees. For these measures, trees were treated as unrooted to allow the stability of the rooting on Epicrionops marmoratus to be assessed (Wilkinson et al. 2002). Relative rates tests were performed using the program RRTree (Robinson et al. 1998). Suboptimal ML trees, conforming to various a priori hypotheses, were found through searches enforcing user-defined topological constraints.

Differences between optimal and suboptimal ML trees were assessed using the Kishino-Hasegawa (KH) test (Kishino & Hasegawa 1989) using RELL with 1000 bootstrap replicates. The KH test is biased when, as here, the trees are not selected a priori because we are more likely to wrongly reject the null hypothesis than we would like at our selected Type I error rate, with the strength of this liberal bias unknown. Thus, whereas failing the test does allow us to accept the null hypothesis, passing the test does not fully justify rejecting the null hypothesis (Goldman et al. 2000). Here a significant result is taken to support the tentative rejection of the null hypothesis, and we used the conservative two-tailed version of the KH test to compensate to some uncertain extent for the liberal bias due to inappropriate tree selection. Although the Shimodaira-Hasegawa test (Shimodaira & Hasegawa 1999) is unbiased, it requires that all plausible trees are included. The identification of this set is problematic for trees with more than only a few taxa, and we have not used this test here.

RESULTS

All PCR amplifications from genomic DNA yielded products of the expected size, which, on sequencing, contained negligible levels of site ambiguity. Some taxa are relatively unstable in the phylogenetic analyses (see below) but there is no obvious reason to suspect that any of the data could have been derived from nuclear copies of mitochondrial sequences.

After incorporation of the new sequences and the exclusion of regions that could not be aligned with confidence, the alignment comprised 914 sites. Of these, 448 were invariant and 123 were parsimony uninformative, leaving 343 parsimony informative sites. There is no significant variation in base composition across the alignment as a whole (χ² tests for homogeneity, \(P = 0.858\), d.f. = 66). In contrast, extensive and significant (\(P = 0.001\)) variation in base composition is evident in the variable sites. Remarkably, if sequences are ranked by their combined GC content, the eight African taxa have the eight highest GC contents (Table 2).
Transition - transversion ratios, based on uncorrected pairwise differences, range from 0.79 to 4.0 (Fig. 1). The many very low ratios occurring in taxa with high total pairwise differences suggest that saturation and/or lineage specific relative rate variation may be a problem in this data set. The four lowest transition - transversion ratios, and nine of the 18 ratios that are less than one, involve the African caciliid Geotrypetes seraphini (Fig. 1), suggesting that saturation or lineage specific rate variation could be a particular problem in accurately placing this taxon. Relative rates tests revealed no significant differences in evolutionary rates in any taxa, with the exceptions that G. seraphini and Typhlonectes natans were both significantly faster than Ichthyophis tricolor (P = 0.045 and P = 0.033, respectively).

The data have a parsimony PTP of 0.01, allowing rejection of the null hypothesis that they contain no more structure than expected by chance alone. Using the likelihood ratio test and the Akaike information criterion, Modeltest selected TrN (Tamura & Nei 1993) + I + G, and GTR (Rodriguez et al. 1990) + I + G models, respectively. We used the simpler TrN + I + G model, and this yielded a single ML tree (Fig. 2).

Relationships among the non-African taxa are mostly those found in previous analyses, with minor differences in the relationships among the Seychellean caeciliids excluding Praslinia, and of the Neotropical caeciliid Siphonops annulatus, both of which were relatively unstable in previous analyses (Wilkinson et al. 2002). Thus there is an Indo-Seychellean caeciliid clade (Gegeneophis, Praslinia, Grandisonia, Hypogeophis), that is more closely related to a Dermophis-Schistometopum clade than to most other caecilians, there is a Typhlonectes-Caecilia clade, and the Uraeotyphlidae and Ichthyophiidae are each other’s closest relatives and sister to all other caecilians except the rhinatrematid Epicrionops. These core relationships were also recovered in parsimony, distance and Bayesian analyses (trees not shown).

Table 2. Base composition and leaf stabilities. Sequences are ranked according to the proportion of guanine and cytosine in the variable sites included in the alignment (GC). Leaf stabilities are from parsimony (MP) and distance (MLD) bootstrap analyses.

<table>
<thead>
<tr>
<th>GC</th>
<th>MP</th>
<th>MLD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Epicrionops marmoratus</td>
<td>0.3231</td>
<td>0.6915</td>
</tr>
<tr>
<td>Ichthyophis tricolor</td>
<td>0.3297</td>
<td>0.7386</td>
</tr>
<tr>
<td>Ichthyophis bannanicus</td>
<td>0.3414</td>
<td>0.7386</td>
</tr>
<tr>
<td>Uraeotyphlus sp.</td>
<td>0.3480</td>
<td>0.738</td>
</tr>
<tr>
<td>Typhlonectes natans</td>
<td>0.3489</td>
<td>0.6271</td>
</tr>
<tr>
<td>Caecilia sp.</td>
<td>0.3575</td>
<td>0.6273</td>
</tr>
<tr>
<td>Siphonops annulatus</td>
<td>0.3616</td>
<td>0.5604</td>
</tr>
<tr>
<td>Hypogeophis rostratus</td>
<td>0.3742</td>
<td>0.7608</td>
</tr>
<tr>
<td>Praslinia cooperi</td>
<td>0.3767</td>
<td>0.8048</td>
</tr>
<tr>
<td>Grandisonia alternans</td>
<td>0.3995</td>
<td>0.7672</td>
</tr>
<tr>
<td>Average</td>
<td>0.4005</td>
<td>0.6949</td>
</tr>
<tr>
<td>Grandisonia brevis</td>
<td>0.4062</td>
<td>0.7778</td>
</tr>
<tr>
<td>Grandisonia larvata</td>
<td>0.4093</td>
<td>0.7643</td>
</tr>
<tr>
<td>Grandisonia seychellensis</td>
<td>0.4100</td>
<td>0.7644</td>
</tr>
<tr>
<td>Gegeneophis ramasawamii</td>
<td>0.4150</td>
<td>0.7604</td>
</tr>
<tr>
<td>Dermophis mexicanus</td>
<td>0.4158</td>
<td>0.7051</td>
</tr>
<tr>
<td>Schistometopum thomense</td>
<td>0.4260</td>
<td>0.7166</td>
</tr>
<tr>
<td>Schistometopum gregorii</td>
<td>0.4318</td>
<td>0.7166</td>
</tr>
<tr>
<td>Boulengerula boulengeri</td>
<td>0.4427</td>
<td>0.6558</td>
</tr>
<tr>
<td>Boulengerula taitanus</td>
<td>0.4439</td>
<td>0.654</td>
</tr>
<tr>
<td>Herpele squalastoma</td>
<td>0.4538</td>
<td>0.6284</td>
</tr>
<tr>
<td>Geotrypetes seraphini</td>
<td>0.4560</td>
<td>0.5476</td>
</tr>
<tr>
<td>Scolecomorphus vittatus</td>
<td>0.4593</td>
<td>0.6182</td>
</tr>
<tr>
<td>Scolecomorphus uluguruensis</td>
<td>0.4820</td>
<td>0.6182</td>
</tr>
</tbody>
</table>
Other interrelationships of African caecilians are generally less well supported. All African caecilians fall within Nussbaum’s (1991) informal ‘higher’ caecilians, a group comprising the Caeciliidae, Typhlonectidae and Scolecomorphidae (united by branch H, Fig. 2). Support for this group is not very strong ($P = 0.67, BP = 60 - 68$) but it is recovered in all optimal trees, is independently supported by morphological phylogenetic analyses (Nussbaum 1979; Duellman & Trueb 1986; Hillis 1991; Wilkinson & Nussbaum 1996; Wilkinson 1997) and is accepted here.

The basal splits within the higher caecilians place the African caeciliids *Boulengerula* and *Herpele squalostoma* together (branch X), and these and the scolecomorphids as successive sister groups of the remaining higher caecilians (branches Y and Z). Each of these relationships has unimpressive bootstrap support and posterior probabilities. Despite lacking strong quantitative support, branch Y is recovered in the optimal trees from each of the analyses employing different methods, and branch X is contradicted only in two of five most parsimonious trees.

The remaining African caeciliid, *G. seraphini*, is nested within a cosmopolitan group of caeciliids (branch C, Fig. 2) that has low bootstrap support and is not recovered in all optimal trees, but which has a surprisingly high posterior probability (0.93). These relationships must be considered speculative and they are accepted only tentatively.

The precise relationships of *G. seraphini* differ greatly in the optimal trees recovered by the different analyses. It is recovered as sister to the *Dermophis-Schistometopum* clade (ML, MLD, Bayesian), sister to the Indo-Seychellian caeciliids (LogDet) or to *Scolecomorphus* (parsimony) and never with strong support. Apart from its tentative inclusion in clade C, all that can be confidently inferred about the relationships of *Geotrypetes* is that it lies outside the *Dermophis-Schistometopum* clade and the Indo-Seychellian clade. Leaf stabilities (Table 2) calculated from parsimony and MLD bootstrap analyses agreed in the rank order, and both identified *G. seraphini* as the least stable taxon. Leaf stabilities for African taxa except *Schistometopum* are lower than average, indicating that their positions are among the relatively least well supported. Leaf stability of *Epicrionops* is close to the average, indicating no special instability in the root.

Constrained analyses produced a number of suboptimal ML trees consistent with various hypotheses of taxonomic, biogeographic and biological interest that were tested against the unconstrained ML tree using the KH test (Table 3). Despite apparent strong support for the monophyly of *Boulengerula*, the best tree in which *Boulengerula* is not monophyletic does

![Figure 1. Scatter plot of pairwise uncorrected estimates of transitions and transversions. The straight line indicates a ratio of 1:1, with points above the line representing particularly low transition - transversion ratios. Shaded points are pairwise estimates of transition transversion ratios of less than one that involve *Geotrypetes seraphini.*](image-url)
not provide a significantly worse fit to the data. In contrast, trees in which the other individual African genera are not monophyletic have a significantly worse fit to the data, as do trees in which African caecilians, African caeciliids, and East and West African caeciliids are monophyletic. Optimal trees in which caecilians with rudimentary eyes are monophyletic can also be tentatively rejected. In contrast, the data do not allow rejection of the hypothesis that viviparity arose only once within caecilians.

### DISCUSSION

Although greatly increasing the taxonomic sampling of African caecilians, our analyses offer incomplete and mostly tentative insights into their phylogenetic relationships. The molecular data support traditional taxonomy in being consistent with the monophyly of the three African genera, *Boulengerula*, *Schistometopum* and *Scolecomorphus* (and of the Scolecomorphidae). *Scolecomorphus* and *Schistometopum* are also characterised by unique morphological synapomorphies (e.g., Nussbaum 1985; Nussbaum & Pfrender 1998; Wake 1998; Gower & Wilkinson 2002; Loader et al. 2003). Nussbaum (1985: 47) reported that “Studies in progress indicate that *Herpele* and *Idiocranium* are distinctive western forms with no close relationship to other African caecilians” whereas our data provide tentative support for the pairing of *Herpele* with *Boulengerula*.

The data also support, albeit weakly, Nussbaum’s (1991) ‘higher’ caecilian clade, as does morphology (Wilkinson & Nussbaum 1996; Wilkinson 1997). The results suggest that the Caeciliidae is paraphyletic, not only with respect to the Typhlonectidae (e.g., Hedges et al. 1993), but also with respect to the Scolecomorphidae, emphasising the need for more comprehensive taxonomic revision. Needless to say, any future revision intended to remove this paraphyly will require greater sampling of caeciliid taxa.

The parallel disjunct distributions of *Schistometopum gregorii* and *Schistometopum thomense*, and of *Scolecomorphus* and *Crotaphatrema* in East and West Africa has been noted previously (e.g., Nussbaum 1985; Nussbaum & Pfrender 1998). The tentative hypothesis that the East African *Boulengerula* and West African *Herpele* are sister taxa, adds a third potential component to this biogeographic parallelism. It is not clear whether the absence of caecilians from Central Africa is real or reflects lack of sampling (Nussbaum & Hinkel 1994), and thus whether the biogeographic pattern is real or apparent. However, this study demonstrates that the caecilian and caeciliid faunas of Africa, and those of East Africa and of West Africa are not monophyletic.

### Table 3. Kishino-Hasegawa tests comparing the fit of the data to the unconstrained ML tree (Fig. 2) and to a range of suboptimal trees, each constrained to make a particular set of taxa monophyletic or not monophyletic. D = difference in log likelihood between optimal and suboptimal trees; P = probability under the null hypothesis that the differences in fit are no greater than expected from random sampling error (noise). 

<table>
<thead>
<tr>
<th>Suboptimal hypothesis</th>
<th>D</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Boulengerula</em> is not monophyletic</td>
<td>9.026</td>
<td>0.086</td>
</tr>
<tr>
<td><em>Schistometopum</em> is not monophyletic</td>
<td>25.229</td>
<td>0.040</td>
</tr>
<tr>
<td><em>Scolecomorphus</em> is not monophyletic</td>
<td>77.902</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>African caecilians are monophyletic</td>
<td>38.623</td>
<td>0.002</td>
</tr>
<tr>
<td>African caeciliids (<em>Boulengerula</em> + <em>Geotrypetes</em> + <em>Herpele</em> + <em>Schistometopum</em>) are monophyletic</td>
<td>37.431</td>
<td>0.006</td>
</tr>
<tr>
<td>East African caeciliids (<em>Boulengerula</em> + <em>Schistometopum gregorii</em>) are monophyletic</td>
<td>68.212</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>West African caeciliids (<em>Herpele</em> + <em>S. thomense</em>) are monophyletic</td>
<td>102.00</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Caeccilians with rudimentary eyes (<em>Boulengerula</em> + <em>Herpele</em> + <em>Gegeneophis</em>) are monophyletic</td>
<td>31.596</td>
<td>0.005</td>
</tr>
<tr>
<td>Viviparous caecilians (<em>Typhlonectes</em> + <em>Dermophis</em> + <em>Geotrypetes</em> + <em>S. thomense</em> + <em>Scolecomorphus</em>) are monophyletic</td>
<td>17.896</td>
<td>0.111</td>
</tr>
</tbody>
</table>
Schistometopum nests within a diverse group of caeciliids as the sister group of Dermophis, with which it was considered congeneric until Parker (1941). In contrast, the Scolecomorpho- idae and Herpele-Boulengerula clades appear to represent relatively deep branches in the higher caecilian clade, with a deep split between the species of Boulengerula as judged by branch lengths. If the split between Schistometopum and Dermophis corresponds to the vicariant separation of Africa from the Neotropics, then this suggests that the origins of some of the current diversity of African caeciliids may predate the break-up of Gondwana.

Boulengerula taitanus was transferred to Afrocaecilia by Taylor (1968) but Afrocaecilia was subsequently synonymised with Boulengerula by Nussbaum & Hinkel (1994) based on phylogenetic analysis of morphological data. Wilkinson et al. (in press) argued that phylogenetic signal from the morphological data is weak and that the synonymy might have been premature. The deep divergence of B. taitanus and B. boulengeri indicated by the molecular data further suggests that a more detailed assessment of the taxonomy of these East African caeciliids is warranted.

Several caecilian genera have closed orbits, and in all of these except the scolecomorphids, the eyes are rudimentary (Wake 1985). It has been argued that rudimentation of the visual system has occurred independently multiple times within caecilians, and that characters of the visual system erroneously group rudimentary-eyed taxa in some morphological phylogenetic analyses (Wilkinson 1997). O’Keefe & Wagner

![Figure 2. Single maximum likelihood tree (LnL = 7832.49). The chosen model of evolution (TrN + I + G) employed a symmetric rate matrix with AG and CT substitutions set at 3.1738 and 8.9975 respectively, and all other substitution types set at unity; base frequencies estimated at 0.4247, 0.2144, 0.1279 and 0.2330 for A, C, G and T respectively; a four category discrete approximation of a gamma distribution (α = 0.57), and the proportion of invariant sites set at 0.276. Numbers in bold are Bayesian posterior probabilities. Numbers in parenthesis are bootstrap proportions from MLD and (where different) parsimony analyses. Letters in bold after taxon names indicate geographic provenance (A = South East Asia, CA = Central America, EA = East Africa, I = India, SA = South America, S = Seychelles, WA = West Africa). Other bold letters indicate internal branches discussed in the text.](image-url)
(2001) further rejected the hypothesis that these characters are evolving independently. Our analysis provides the first support from molecular data for the parallel rudimentation of visual systems in caecilians, with this occurring independently in at least two lineages - *Boulengerula* (+ *Herpele*) and *Gegeneophis*.

Our analyses also suggest that viviparity has evolved at least three times, but using the KH test we are unable to reject trees in which all viviparous caecilians are a monophyletic group. The trees for this test were obtained using a backbone constraint that excluded *Schistometopum gregorii* and *Herpele squalostoma* because the reproductive modes of these species are uncertain (Wilkinson & Nussbaum 1998). Natives informed Loveridge (1936) that *S. gregorii* laid eggs in water, but we concur with Nussbaum & Pfrender (1998) that this is unlikely. The placement of *S. gregorii* and *H. squalostoma* in the optimal trees suggests that they are viviparous and oviparous respectively, predictions that can be tested empirically.

Two caeciliids, the West African *Geotrypetes seraphini* and the South American *Siphonops annulatus* are particularly unstable. Their phylogenetic placement is sensitive to method of analysis, is never well supported, and they have the lowest leaf stabilities. Little can be said of their relationships, other than that they lie somewhere within the higher caecilian clade, probably not basally, and that they lie outside the well-supported groupings of the scolecomorphids, *Typhlonectes* + *Caecilia*, *Dermophis* + *Schistometopum* or the Indo-Seychellenian clade.

Although partial sequences of mitochondrial 16S and 12S have provided important insights into the phylogeny of caecilians (Hedges *et al.* 1993; Gower *et al.* 2002; Wilkinson *et al.* 2002), they are not adequate for resolving the relationships of African caecilians. In addition, there are strong base compositional biases and some very low transition - transversion ratios that suggest saturation and/or lineage specific rate variation may hinder accurate phylogenetic inference from these data. These potential problems seem particularly to affect comparisons involving the African caecilian sequences, all of which have below average leaf stabilities (Table 2). African sequences appear to provide disproportionately low estimates of transition - transversion ratios and this is particularly true of *Geotrypetes seraphini*. It is also remarkable that, when taxa are ranked by their GC content, the African caecilians exclusively occupy the highest ranks (Table 2). We would not have predicted any simple correlation between geography and base composition, are unaware of any comparable patterns in the literature, and consider its existence intriguing and worthy of further study.

Given that our alignment includes only 16 of 33 genera and 23 of c.160 caecilian species, partial 16S and 12S data should continue to be useful for placing many of the currently unsampled taxa. However, better resolution of the relationships of the African caecilians is unlikely to be achieved purely through the benefits of denser taxonomic sampling and will probably require data from more genes and morphology.

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LITERATURE CITED


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